

REVIEW

Open Access



Recent developments on the role of epigenetics in obesity and metabolic disease

Susan J. van Dijk¹, Ross L. Tellam², Janna L. Morrison³, Beverly S. Muhlhauser^{4,5†} and Peter L. Molloy^{1*†}

Abstract

The increased prevalence of obesity and related comorbidities is a major public health problem. While genetic factors undoubtedly play a role in determining individual susceptibility to weight gain and obesity, the identified genetic variants only explain part of the variation. This has led to growing interest in understanding the potential role of epigenetics as a mediator of gene-environment interactions underlying the development of obesity and its associated comorbidities. Initial evidence in support of a role of epigenetics in obesity and type 2 diabetes mellitus (T2DM) was mainly provided by animal studies, which reported epigenetic changes in key metabolically important tissues following high-fat feeding and epigenetic differences between lean and obese animals and by human studies which showed epigenetic changes in obesity and T2DM candidate genes in obese/diabetic individuals. More recently, advances in epigenetic methodologies and the reduced cost of epigenome-wide association studies (EWAS) have led to a rapid expansion of studies in human populations. These studies have also reported epigenetic differences between obese/T2DM adults and healthy controls and epigenetic changes in association with nutritional, weight loss, and exercise interventions. There is also increasing evidence from both human and animal studies that the relationship between perinatal nutritional exposures and later risk of obesity and T2DM may be mediated by epigenetic changes in the offspring. The aim of this review is to summarize the most recent developments in this rapidly moving field, with a particular focus on human EWAS and studies investigating the impact of nutritional and lifestyle factors (both pre- and postnatal) on the epigenome and their relationship to metabolic health outcomes. The difficulties in distinguishing consequence from causality in these studies and the critical role of animal models for testing causal relationships and providing insight into underlying mechanisms are also addressed. In summary, the area of epigenetics and metabolic health has seen rapid developments in a short space of time. While the outcomes to date are promising, studies are ongoing, and the next decade promises to be a time of productive research into the complex interactions between the genome, epigenome, and environment as they relate to metabolic disease.

Keywords: Epigenetics, DNA methylation, Obesity, Type 2 diabetes, Developmental programming

Introduction

Obesity is a complex, multifactorial disease, and better understanding of the mechanisms underlying the interactions between lifestyle, environment, and genetics is critical for developing effective strategies for prevention and treatment [1].

In a society where energy-dense food is plentiful and the need for physical activity is low, there is a wide variation in individuals' susceptibility to develop

obesity and metabolic health problems. Estimates of the role of heredity in this variation are in the range of 40–70 %, and while large genome-wide association studies (GWAS) have identified a number of genetic loci associated with obesity risk, the ~100 most common genetic variants only account for a few percent of variance in obesity [2, 3]. Genome-wide estimates are higher, accounting for ~20 % of the variation [3]; however, a large portion of the heritability remains unexplained.

Recently, attention has turned to investigating the role of epigenetic changes in the etiology of obesity. It has been argued that the epigenome may represent the mechanistic link between genetic variants and environmental

* Correspondence: Peter.Molloy@csiro.au

†Equal contributors

¹CSIRO Food and Nutrition Flagship, PO Box 52, North Ryde, NSW 1670, Australia

Full list of author information is available at the end of the article

factors in determining obesity risk and could help explain the “missing heritability.” The first human epigenetic studies were small and only investigated a limited number of loci. While this generally resulted in poor reproducibility, some of these early findings, for instance the relationship between *PGC1A* methylation and type 2 diabetes mellitus (T2DM) [4] and others as discussed in van Dijk et al. [5], have been replicated in later studies. Recent advances and increased affordability of high-throughput technologies now allow for large-scale epigenome-wide association studies (EWAS) and integration of different layers of genomic information to explore the complex interactions between the genotype, epigenome, transcriptome, and the environment [6–9]. These studies are still in their infancy, but the results thus far have shown promise in helping to explain the variation in obesity susceptibility.

There is increasing evidence that obesity has developmental origins, as exposure to a suboptimal nutrient supply before birth or in early infancy is associated with an increased risk of obesity and metabolic disease in later life [10–13]. Initially, animal studies demonstrated that a range of early life nutritional exposures, especially those experienced early in gestation, could induce epigenetic changes in key metabolic tissues of the offspring that persisted after birth and result in permanent alterations in gene function [13–17]. Evidence is emerging to support the existence of the same mechanism in humans. This has led to a search for epigenetic marks present early in life that predict later risk of metabolic disease, and studies to determine whether epigenetic programming of metabolic disease could be prevented or reversed in later life.

This review provides an update of our previous systematic review of studies on epigenetics and obesity in humans [5]. Our previous review showcased the promising outcomes of initial studies, including the first potential epigenetic marks for obesity that could be detected at birth (e.g., *RXR α*) [18]. However, it also highlighted the limited reproducibility of the findings and the lack of larger scale longitudinal investigations. The current review focuses on recent developments in this rapidly moving field and, in particular, on human EWAS and studies investigating the impact of (pre- and postnatal) nutritional and lifestyle factors on the epigenome and the emerging role of epigenetics in the pathology of obesity. We also address the difficulties in identifying causality in these studies and the importance of animal models in providing insight into mechanisms.

Review

Epigenetic changes in animal models of obesity

Animal models provide unique opportunities for highly controlled studies that provide mechanistic insight into

the role of specific epigenetic marks, both as indicators of current metabolic status and as predictors of the future risk of obesity and metabolic disease. A particularly important aspect of animal studies is that they allow for the assessment of epigenetic changes within target tissues, including the liver and hypothalamus, which is much more difficult in humans. Moreover, the ability to harvest large quantities of fresh tissue makes it possible to assess multiple chromatin marks as well as DNA methylation. Some of these epigenetic modifications either alone or in combination may be responsive to environmental programming. In animal models, it is also possible to study multiple generations of offspring and thus enable differentiation between transgenerational and intergenerational transmission of obesity risk mediated by epigenetic memory of parental nutritional status, which cannot be easily distinguished in human studies. We use the former term for meiotic transmission of risk in the absence of continued exposure while the latter primarily entails direct transmission of risk through metabolic reprogramming of the fetus or gametes.

Animal studies have played a critical role in our current understanding of the role of epigenetics in the developmental origins of obesity and T2DM. Both increased and decreased maternal nutrition during pregnancy have been associated with increased fat deposition in offspring of most mammalian species studied to date (reviewed in [11, 13–15, 19]). Maternal nutrition during pregnancy not only has potential for direct effects on the fetus, it also may directly impact the developing oocytes of female fetuses and primordial germ cells of male fetuses and therefore could impact both the offspring and grand-offspring. Hence, multigenerational data are usually required to differentiate between maternal intergenerational and transgenerational transmission mechanisms.

Table 1 summarizes a variety of animal models that have been used to provide evidence of metabolic and epigenetic changes in offspring associated with the parental plane of nutrition. It also contains information pertaining to studies identifying altered epigenetic marks in adult individuals who undergo direct nutritional challenges. The table is structured by suggested risk transmission type.

- (i) Epigenetic changes in offspring associated with maternal nutrition during gestation

Maternal nutritional supplementation, undernutrition, and overnutrition during pregnancy can alter fat deposition and energy homeostasis in offspring [11, 13–15, 19]. Associated with these effects in the offspring are changes in DNA methylation, histone post-translational modifications, and gene expression for several target genes,

Table 1 Evidence for a role of epigenetics in animal models of obesity separated by transmission type

Transmission type	Species	Experimental model	Phenotype affected	Epigenetic changes	Ref.
Intergenerational maternal effect	Sheep	Periconceptional undernutrition in normal and overweight ewes using artificial insemination and embryo transfer	Fat deposition and adrenal changes in offspring	Decreased expression of <i>IGF2</i> and decreased DNA methylation of a proximal imprinting control region; changes in adrenal <i>IGF2</i> DNA methylation; hypermethylation of pituitary glucocorticoid receptor	[14, 88–91]
Intergenerational maternal effect	Sheep	Maternal undernutrition prior to conception and during early gestation	Programming of obesity	Altered offspring histone methylation and acetylation in fetal hypothalamic energy regulating pathways	[20]
Intergenerational maternal effect	Sheep	Different maternal dietary energy sources during that last half of gestation	Late gestation fetal gene expression and DNA methylation from a variety of tissues	Changes in late gestation fetal DNA methylation of CpG islands associated with <i>IGF2R</i> and <i>H19</i> in muscle and adipose tissue	[23]
Intergenerational maternal effect	Pig	Methylating micronutrient supplementation during gestation—impacts on F2	Back fat percentage, adipose tissue, and fat thickness at 10th rib, croup, and shoulder in F2	Differentially expressed metabolic genes in F2 liver and muscle, DNA methylation change in <i>IGF1</i>	[31]
Intergenerational maternal effect	Mouse	Maternal low-protein diet during gestation and maternal diet restriction during gestation	Body weight, food intake, and adiposity	Altered germline DNA methylation of F1 adult males in a locus specific manner; changed expression and DNA methylation of <i>LXRα</i> in liver; demethylation of leptin promoter in adipocytes	[22, 26, 92]
Intergenerational maternal effect	Mouse	Maternal high-fat diet during gestation; maternal obesity model; maternal high-fat diet using a <i>Glut4^{+/-}</i> genetic background; maternal diet-induced obesity	Offspring chromatin organization; metabolic syndrome in offspring unmasked by exposure to western diet; glucose intolerance, insulin resistance, hepatic steatosis; obesity; exacerbated metabolic syndrome in offspring; insulin levels, insulin resistance in adipose tissue	Changes in offspring hepatic histone marks H3K14ac and H3K9me3; changes in offspring hepatic gene expression and widespread subtle changes in cytosine methylation; DNA methylation change in <i>PEG3</i> in spermatozoa of offspring; cell autonomous transmission of altered insulin signaling. Reduced <i>IRS1</i> expression associated with elevated miR126	[21, 24, 28, 93]
Intergenerational maternal effect	Rat	Maternal diet restriction during gestation; suboptimal diet during early gestation	Catch up growth, obesity, and liver weight; T2D	Change in offspring liver <i>IGF1</i> expression and <i>IGF1</i> H3K4 methylation; decreased <i>PPARA</i> expression and increased DNA methylation in the <i>PPARA</i> promoter in liver; change in offspring growth hormone and <i>PPARA</i> expression and DNA methylation in liver; chromatin changes affecting enhancer/promoter interactions at <i>HNF4A</i> promoter in pancreatic islets from offspring	[17, 25, 29, 30]
Intergenerational maternal effect	Rat	Maternal overfeeding model during preconception and gestation	Adipogenesis, gene expression and reduced representation DNA methylation in offspring	Changes in gene expression and proximal DNA methylation in genes in lipogenic pathways of adipocytes from offspring	[16]
Intergenerational maternal effect	Macaque	Maternal high-fat diet during gestation	Altered expression of <i>Npas2</i>	Changes in offspring fetal liver chromatin mark H3K14ac in the <i>NPAS2</i> promoter	[27]
Intergenerational paternal effect	Drosophila	Paternal overnutrition	Obesity in offspring	Chromatin (H3K9me3 and H3K27me3)-dependent reprogramming of offspring metabolic genes; a similar system may regulate obesity susceptibility and phenotypic variation in mice and humans	[33]
Intergenerational paternal effect	Mouse	Paternal low-protein diet	High cholesterol in offspring	Changes in hepatic gene expression and DNA methylation in offspring	[32]

Table 1 Evidence for a role of epigenetics in animal models of obesity separated by transmission type (*Continued*)

Intergenerational paternal effect	Mouse	Intrauterine growth restriction	F1 offspring become obese and glucose intolerant with aging	F1 males show change in methylation of <i>LXRA</i> in sperm that is transmitted to somatic cells in the F2	[34]
Intergenerational paternal effect	Mouse	Paternal prediabetes	F1 has increased susceptibility to diabetes	F1 show changes in pancreatic gene expression and DNA methylation linked to insulin signaling. A large portion of these genes are also differentially methylated in sperm	[35]
Potential transgenerational effect	Mouse	A ^{vy} mouse—change in coat color and adult onset obesity through maternal transmission to the next generation; modulation by methyl donors and genistein during gestation.	Coat color and adult onset obesity in offspring	DNA methylation of a retrotransposon promoter adjacent to the <i>agouti</i> gene; evidence for germ line transmission of methylation status	[38–41, 94]
Effect resulting from direct exposure of adult	Mouse	High-fat diet	Weight, fasting glucose, glucose, and insulin tolerance tests; obesity	Differential DNA methylation at numerous sites in adipose tissue; changes in DNA methylation of metabolism-related genes in liver and oocytes	[43, 95]

T2D type 2 diabetes

especially genes regulating fatty acid metabolism and insulin signaling [16, 17, 20–30]. The diversity of animal models used in these studies and the common metabolic pathways impacted suggest an evolutionarily conserved adaptive response mediated by epigenetic modification. However, few of the specific identified genes and epigenetic changes have been cross-validated in related studies, and large-scale genome-wide investigations have typically not been applied. A major hindrance to comparison of these studies is the different developmental windows subjected to nutritional challenge, which may cause considerably different outcomes. Proof that the epigenetic changes are causal rather than being associated with offspring phenotypic changes is also required. This will necessitate the identification of a parental nutritionally induced epigenetic “memory” response that precedes development of the altered phenotype in offspring.

(ii) Effects of paternal nutrition on offspring epigenetic marks

Emerging studies have demonstrated that paternal plane of nutrition can impact offspring fat deposition and epigenetic marks [31–34]. One recent investigation using mice has demonstrated that paternal prediabetes leads to increased susceptibility to diabetes in F1 offspring with associated changes in pancreatic gene expression and DNA methylation linked to insulin signaling [35]. Importantly, there was an overlap of these epigenetic changes in pancreatic islets and sperm suggesting germ line inheritance. However, most of these studies, although intriguing in their implications, are limited in the genomic scale of investigation and frequently show weak and somewhat transient epigenetic alterations associated with mild metabolic phenotypes in offspring.

(iii) Potential transgenerational epigenetic changes promoting fat deposition in offspring

Stable transmission of epigenetic information across multiple generations is well described in plant systems and *C. elegans*, but its significance in mammals is still much debated [36, 37]. An epigenetic basis for grandparental transmission of phenotypes in response to dietary exposures has been well established, including in livestock species [31]. The most influential studies demonstrating effects of epigenetic transmission impacting offspring phenotype have used the example of the viable yellow agouti (A^{vy}) mouse [38]. In this mouse, an insertion of a retrotransposon upstream of the *agouti* gene causes its constitutive expression and consequent yellow coat color and adult onset obesity. Maternal transmission through the germ line results in DNA methylation

mediated silencing of *agouti* expression resulting in wild-type coat color and lean phenotype of the offspring [39, 40]. Importantly, subsequent studies in these mice demonstrated that maternal exposure to methyl donors causes a shift in coat color [41]. One study has reported transmission of a phenotype to the F3 generation and alterations in expression of large number of genes in response to protein restriction in F0 [42]; however, alterations in expression were highly variable and a direct link to epigenetic changes was not identified in this system.

(iv) Direct exposure of individuals to excess nutrition in postnatal life

While many studies have identified diet-associated epigenetic changes in animal models using candidate site-specific regions, there have been few genome-wide analyses undertaken. A recent study focussed on determining the direct epigenetic impact of high-fat diets/diet-induced obesity in adult mice using genome-wide gene expression and DNA methylation analyses [43]. This study identified 232 differentially methylated regions (DMRs) in adipocytes from control and high-fat fed mice. Importantly, the corresponding human regions for the murine DMRs were also differentially methylated in adipose tissue from a population of obese and lean humans, thereby highlighting the remarkable evolutionary conservation of these regions. This result emphasizes the likely importance of the identified DMRs in regulating energy homeostasis in mammals.

Human studies

Drawing on the evidence from animal studies and with the increasing availability of affordable tools for genome-wide analysis, there has been a rapid expansion of epigenome studies in humans. These studies have mostly focused on the identification of site-specific differences in DNA methylation that are associated with metabolic phenotypes.

A key question is the extent to which epigenetic modifications contribute to the development of the metabolic phenotype, rather than simply being a consequence of it (Fig. 1). Epigenetic programming could contribute to obesity development, as well as playing a role in consequent risk of cardiovascular and metabolic problems. In human studies, it is difficult to prove causality [44], but inferences can be made from a number of lines of evidence:

- (i) Genetic association studies. Genetic polymorphisms that are associated with an increased risk of developing particular conditions are a priori linked to the causative genes. The presence of differential

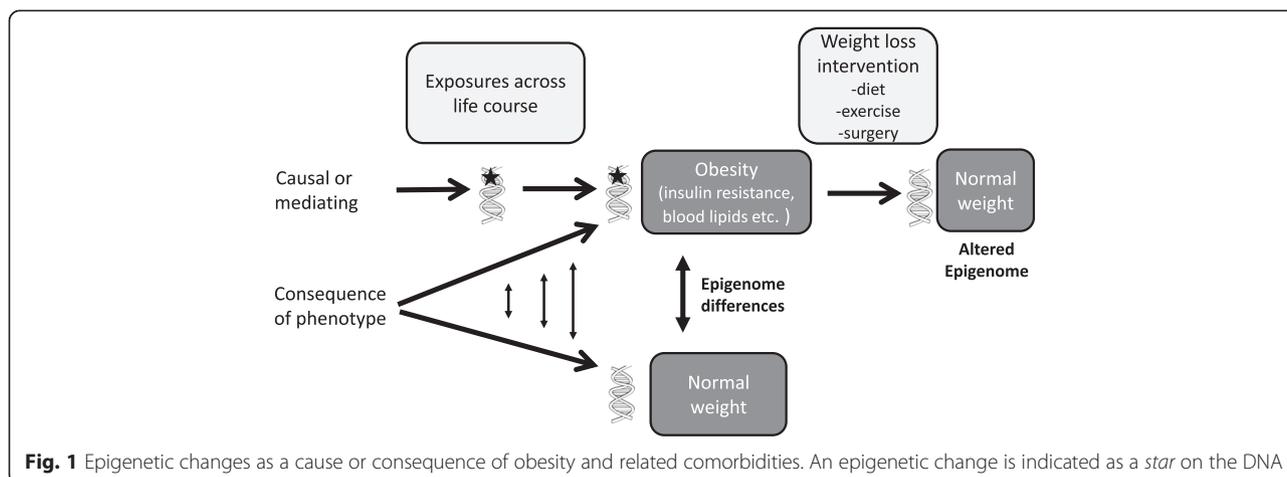


Fig. 1 Epigenetic changes as a cause or consequence of obesity and related comorbidities. An epigenetic change is indicated as a star on the DNA

methylation in such regions infers functional relevance of these epigenetic changes in controlling expression of the proximal gene(s). There are strong cis-acting genetic effects underpinning much epigenetic variation [7, 45], and in population-based studies, methods that use genetic surrogates to infer a causal or mediating role of epigenome differences have been applied [7, 46–48]. The use of familial genetic information can also lead to the identification of potentially causative candidate regions showing phenotype-related differential methylation [49].

- (ii) Timing of epigenetic changes. The presence of an epigenetic mark prior to development of a phenotype is an essential feature associated with causality. Conversely, the presence of a mark in association with obesity, but not before its development, can be used to exclude causality but would not exclude a possible role in subsequent obesity-related pathology.
- (iii) Plausible inference of mechanism. This refers to epigenetic changes that are associated with altered expression of genes with an established role in regulating the phenotype of interest. One such example is the association of methylation at two CpG sites at the *CPT1A* gene with circulating triglyceride levels [50]. *CPT1A* encodes carnitine palmitoyltransferase 1A, an enzyme with a central role in fatty acid metabolism, and this is strongly indicative that differential methylation of this gene may be causally related to the alterations in plasma triglyceride concentrations.

Epigenome-wide association studies: identifying epigenetic biomarkers of metabolic health

A number of recent investigations have focused on exploring associations between obesity/metabolic diseases

and DNA methylation across the genome (Table 2). The largest published EWAS so far, including a total of 5465 individuals, identified 37 methylation sites in blood that were associated with body mass index (BMI), including sites in *CPT1A*, *ABCG1*, and *SREBF1* [51]. Another large-scale study showed consistent associations between BMI and methylation in *HIF3A* in whole blood and adipose tissue [52], a finding which was also partially replicated in other studies [9, 51]. Other recently reported associations between obesity-related measures and DNA methylation include (i) DNA methylation differences between lean and obese individuals in *LY86* in blood leukocytes [53]; (ii) associations between *PGC1A* promoter methylation in whole blood of children and adiposity 5 years later [54]; (iii) associations between waist-hip ratio and *ADRB3* methylation in blood [55]; and (iv) associations between BMI, body fat distribution measures, and multiple DNA methylation sites in adipose tissue [9, 56]. EWAS have also shown associations between DNA methylation sites and blood lipids [55, 57–59], serum metabolites [60], insulin resistance [9, 61], and T2DM [48, 62, 63] (Table 2).

From these studies, altered methylation of *PGC1A*, *HIF3A*, *ABCG1*, and *CPT1A* and the previously described *RXRA* [18] have emerged as biomarkers associated with, or perhaps predictive of, metabolic health that are also plausible candidates for a role in development of metabolic disease.

Interaction between genotype and the epigenome

Epigenetic variation is highly influenced by the underlying genetic variation, with genotype estimated to explain ~20–40 % of the variation [6, 8]. Recently, a number of studies have begun to integrate methylome and genotype data to identify methylation quantitative trait loci (meQTL) associated with disease phenotypes. For instance, in adipose tissue, an meQTL overlapping

Table 2 Representative human studies showing evidence for a role of epigenetics in obesity and related comorbidities

Phenotype	Association with epigenetic marks or changes in epigenetic marks ^a	Includes validation ^b	Ref.
Cross-sectional studies			
BMI, WC	Methylation in 37 CpGs associated with BMI and 1 probe with WC in blood ($n = 5465$); of those 16 CpGs (e.g., CpGs in <i>CPT1A</i> , <i>ABCG1</i> , and <i>SREBF1</i>) were also associated with BMI in subcutaneous adipose tissue ($n = 648$)	Yes, 3 other cohorts	[51]
BMI	DNA methylation of 4979 CpGs (e.g., in <i>FTO</i> , <i>TCF7L2</i> , <i>FASN</i> , <i>PGC1A</i> , <i>CCRL2</i>) in subcutaneous adipose tissue ($n = 190$). Several BMI-related methylation sites were also associated with age (e.g., <i>ELOVL2</i>), and with gene expression levels	Yes, 2nd cohort	[9]
BMI	<i>HIF3A</i> methylation 3 CpGs in whole blood ($n = 2587$) and subcutaneous adipose tissue ($n = 635$), not in skin ($n = 395$)	Yes, 3 other cohorts	[52]
Obesity	<i>LY86</i> methylation 1 CpG in blood leukocytes adolescents and adults ($n = 1534$)	Yes, 4 other cohorts	[53]
Obesity	249 DMRs in subcutaneous adipose tissue ($n = 21$), methylation in all these 249 regions also changed with high-fat feeding in mice and includes regions overlapping T2DM loci such as in <i>TCF7L2</i>	No, only in mice	[43]
WHR, blood pressure LDL cholesterol	Association between WHR and <i>ADRB3</i> methylation in whole blood and between blood pressure and <i>ADRB3</i> methylation in visceral adipose tissue of obese ($n = 25$). Association between <i>ADRB3</i> methylation and LDL cholesterol in whole blood of men with familial hypercholesterolemia ($n = 41$)	Not validated in 2nd cohort	[55]
BMI	<i>ADCY3</i> meQTL in subcutaneous adipose tissue from twins ($n = 648$)	Yes, technical	[8]
BMI	Methylation differences at 1236 CpGs in leukocytes of monozygotic twins discordant for BMI and liver fat ($n = 13$)	Yes, technical	[96]
Adiposity phenotypes	Methylation in 101 genes in subcutaneous adipose tissue ($n = 106$) including <i>AOC3</i> , <i>SOD3</i> , <i>DOCK9</i> , <i>AQP7</i> , <i>ANGPT4</i> , <i>ANGPT2</i> , <i>TIMP4</i> , <i>ADAMST4</i> , <i>HOXA3</i> , and <i>LIPE</i> , not found in blood leukocytes same individuals	No validation performed	[56]
TG, VLDL	<i>CPT1A</i> methylation 1 CpG in CD4+T cells ($n = 991$) and leukocytes ($n = 1261$)	Yes, technical and 2nd cohort	[57]
VLDL and LDL	<i>CPT1A</i> methylation 2 CpGs in CD4+T cells ($n = 994$)	Yes, in the same cohort	[58]
Cholesterol and TG	Methylation in 9 CpGs, including in <i>ABCG1</i> , <i>CPT1A</i> and <i>SREBF1</i> , in whole blood ($n = 2747$), 5 of these CpGs also showed associations in subcutaneous adipose tissue ($n = 634$)	Yes, 2 other cohorts	[59]
Insulin and HOMA-IR	<i>ABCG1</i> 1 CpG in CD4+T cells ($n = 837$)	Yes, in the same cohort	[61]
HbA1C	DNA methylation of 711 CpGs in subcutaneous adipose tissue ($n = 96$ males). Not validated in female cohort ($n = 94$) but 30 CpGs validated in T2DM case-control cohort	Yes, validated in 1 of 2 cohorts	[9]
T2DM	<i>MALT1</i> methylation whole blood ($n = 27$ twins + $n = 263$ individuals) and other less significant DMRs (FDR < 0.1) overlapping T2DM GWAS loci	Yes, other cohort	[48]
T2DM	Methylation in 1649 CpG sites, some overlapping T2DM, and obesity GWAS loci such as <i>TCF7L2</i> , <i>FTO</i> , and <i>KCNQ1</i> in pancreatic islets ($n = 49$)	Yes, technical	[62]
T2DM	No differentially methylated sites (after FDR correction) in T2DM discordant monozygotic twins ($n = 14$ pairs). Differential methylation at 15,627 CpGs, including in T2DM GWAS loci <i>PPARG</i> , <i>KCNQ1</i> , <i>TCF7L2</i> , and <i>IRS1</i> in subcutaneous adipose tissue ($n = 56$). 1410 of these CpGs were also differentially methylated ($P < 0.05$) in the T2DM discordant twins	Yes, other cohort	[63]

Table 2 Representative human studies showing evidence for a role of epigenetics in obesity and related comorbidities (*Continued*)

Longitudinal studies			
Adiposity measured annually age 9–14 years	Increase <i>PGC1A</i> promoter methylation in whole blood children measured annually from 5–7 years (<i>n</i> = 40)	No, but measures at multiple time points	[54]
Maternal exposure or phenotype and epigenetic marks in offspring			
Prenatal famine	181 DMRs in adult whole blood (<i>n</i> = 48), including in <i>CDH23</i> , <i>SMAD7</i> , <i>INSR</i> , <i>CPT1A</i> , <i>KLF13</i> , <i>RFTN1</i> (validated in <i>n</i> = 120)	Yes, technical and 2nd cohort	[68]
Variation methyl donor intake	Changes in mean methylation across <i>BOLA3</i> , <i>LOC654433</i> , <i>EXD3</i> , <i>ZFYVE28</i> , <i>RBM46</i> , and <i>ZNF678</i> in blood leukocytes (<i>n</i> = 126) and hair follicles (<i>n</i> = 82) of 2- to 8-month infants	No validation performed	[69]
Periconceptional BMI	Decreased mean methylation across <i>BOLA3</i> , <i>LOC654433</i> , <i>EXD3</i> , <i>ZFYVE28</i> , <i>RBM46</i> , and <i>ZNF678</i> in blood leukocytes (<i>n</i> = 126) and hair follicles (<i>n</i> = 82) of 2- to 8-month infants	No validation performed	[69]
Gestational weight gain early pregnancy	Increased methylation 4 CpGs in <i>MMP7</i> , <i>KCNK4</i> , <i>TRPM5</i> , and <i>NFKB1</i> in newborn cord blood (<i>n</i> = 88), no association in 2nd cohort (<i>n</i> = 170)	Not validated (technical and 2nd cohort)	[71]
Preconceptional BMI	Differential methylation in <i>ZCCHC10</i> in newborn cord blood (<i>n</i> = 308), other less significant sites were found in <i>WNT16</i> , <i>ACPL2</i> , <i>C18orf8</i> , <i>ANGPTL2</i> , <i>SAPCD2</i> , and <i>ADCY3</i>	No	[73]
Gestational Diabetes	42 CpGs in newborn cord blood (<i>n</i> = 136). ~1/3 of CpGs overlapped with sites that were associated with maternal glucose levels (<i>n</i> = 36) or micronutrient supplementation (<i>n</i> = 59) in 2 other child cohorts	Yes, technical and 2 other cohorts	[70]
Gestational Diabetes	No differentially methylated sites (after FDR correction) in cord blood and placenta (<i>n</i> = 44) but enrichment of sites in genes metabolic disease pathway	No	[72]
Intervention			
Weight loss surgery	Change in methylation at 3601 CpGs (195 DMRs) in subcutaneous adipose tissue and 15 CpGs in omental adipose tissue (<i>n</i> = 15), some DMRs overlapping known obesity and T2DM loci	Yes, technical	[64]
Weight loss surgery	227 DMRs, methylation in these regions also changed with high-fat feeding in mice	No, only in mice	[43]
Weight loss surgery in liver disease	Before surgery 467 differentially methylated CpGs between control (<i>n</i> = 18), healthy obese (<i>n</i> = 18), steatosis (<i>n</i> = 12), and NASH (<i>n</i> = 15) liver samples. After surgery changes in methylation at 113 CpGs, disease-associated methylation was reversible at the <i>HOXB1</i> , <i>PRKCZ</i> , <i>SLC38A10</i> , and <i>SECTM1</i> loci	No, but for baseline technical and 2nd cohort	[79]
Weight loss	Methylation profiles of <i>RYR1</i> , <i>TUBA3C</i> and <i>BDNF</i> in PBMCs of successful weight loss maintainers (<i>n</i> = 16) more closely resembled lean (<i>n</i> = 16) than obese subjects (<i>n</i> = 16)	No	[77]
Endurance and strength exercise	Changes in DNA methylation in skeletal muscle of obese T2DM subjects (<i>n</i> = 17) after 16 weeks, most pronounced with endurance exercise	No	[83]
High-fat diet (5 days)	<i>PPARGC1A</i> DNA methylation across 4 CpGs increased in subcutaneous adipose tissue of lean adults born with low birth weight (<i>n</i> = 19) but not in controls (<i>n</i> = 26)	No	[76]

BMI body mass index, *DMR* differentially methylated region, *GDM* gestational diabetes mellitus, *GWAS* genome-wide association study, *NASH* non-alcoholic steatohepatitis, *PBMC* peripheral blood mononuclear cell, *T2DM* type 2 diabetes mellitus, *TG* triglycerides, *WC* waist circumference, *WHR* waist-hip ratio

^aIf not otherwise stated the number of subjects includes the total number in discovery and validation sets

^bValidation refers to validation of at least some epigenetic marks identified in the primary study cohort. This can be either technical validation, using another method for the measurement of epigenetic marks, or validation in another cohort

with a BMI genetic risk locus has been identified in an enhancer element upstream of *ADCY3* [8]. Other studies have also identified overlaps between known obesity and T2DM risk loci and DMRs associated with obesity and T2DM [43, 48, 62]. Methylation of a number of such DMRs was also modulated by high-fat feeding in mice [43] and weight loss in humans [64]. These results identify an intriguing link between genetic variations linked with disease susceptibility and their association with regions of the genome that undergo epigenetic modifications in response to nutritional challenges, implying a causal relationship. The close connection between genetic and epigenetic variation may signify their essential roles in generating individual variation [65, 66]. However, while these findings suggest that DNA methylation may be a mediator of genetic effects, it is also important to consider that both genetic and epigenetic processes could act independently on the same genes. Twin studies [8, 63, 67] can provide important insights and indicate that inter-individual differences in levels of DNA methylation arise predominantly from non-shared environment and stochastic influences, minimally from shared environmental effects, but also with a significant impact of genetic variation.

The impact of the prenatal and postnatal environment on the epigenome

Prenatal environment Two recently published studies made use of human populations that experienced “natural” variations in nutrient supply to study the impact of maternal nutrition before or during pregnancy on DNA methylation in the offspring [68, 69]. The first study used a Gambian mother-child cohort to show that both seasonal variations in maternal methyl donor intake during pregnancy and maternal pre-pregnancy BMI were associated with altered methylation in the infants [69]. The second study utilized adult offspring from the Dutch Hunger Winter cohort to investigate the effect of prenatal exposure to an acute period of severe maternal undernutrition on DNA methylation of genes involved in growth and metabolism in adulthood [68]. The results highlighted the importance of the timing of the exposure in its impact on the epigenome, since significant epigenetic effects were only identified in individuals exposed to famine during early gestation. Importantly, the epigenetic changes occurred in conjunction with increased BMI; however, it was not possible to establish in this study whether these changes were present earlier in life or a consequence of the higher BMI.

Other recent studies have provided evidence that prenatal overnutrition and an obese or diabetic maternal environment are also associated with DNA methylation changes in genes related to embryonic development, growth, and metabolic disease in the offspring [70–73].

While human data are scarce, there are indications that paternal obesity can lead to altered methylation of imprinted genes in the newborn [74], an effect thought to be mediated via epigenetic changes acquired during spermatogenesis.

Postnatal environment The epigenome is established de novo during embryonic development, and therefore, the prenatal environment most likely has the most significant impact on the epigenome. However, it is now clear that changes do occur in the “mature” epigenome under the influence of a range of conditions, including aging, exposure to toxins, and dietary alterations. For example, changes in DNA methylation in numerous genes in skeletal muscle and *PGC1A* in adipose tissue have been demonstrated in response to a high-fat diet [75, 76]. Interventions to lose body fat mass have also been associated with changes in DNA methylation. Studies have reported that the DNA methylation profiles of adipose tissue [43, 64], peripheral blood mononuclear cells [77], and muscle tissue [78] in formerly obese patients become more similar to the profiles of lean subjects following weight loss. Weight loss surgery also partially reversed non-alcoholic fatty liver disease-associated methylation changes in liver [79] and in another study led to hypomethylation of multiple obesity candidate genes, with more pronounced effects in subcutaneous compared to omental (visceral) fat [64]. Accumulating evidence suggests that exercise interventions can also influence DNA methylation. Most of these studies have been conducted in lean individuals [80–82], but one exercise study in obese T2DM subjects also demonstrated changes in DNA methylation, including in genes involved in fatty acid and glucose transport [83]. Epigenetic changes also occur with aging, and recent data suggest a role of obesity in augmenting them [9, 84, 85]. Obesity accelerated the epigenetic age of liver tissue, but in contrast to the findings described above, this effect was not reversible after weight loss [84].

Collectively, the evidence in support of the capacity to modulate the epigenome in adults suggests that there may be the potential to intervene in postnatal life to modulate or reverse adverse epigenetic programming.

Effect sizes and differences between tissue types

DNA methylation changes associated with obesity or induced by diet or lifestyle interventions and weight loss are generally modest (<15 %), although this varies depending on the phenotype and tissue studied. For instance, changes greater than 20 % have been reported in adipose tissue after weight loss [64] and associations between *HIF3A* methylation and BMI in adipose tissue were more pronounced than in blood [52].

The biological relevance of relatively small methylation changes has been questioned. However, in tissues consisting of a mixture of cell types, a small change in DNA methylation may actually reflect a significant change in a specific cell fraction. Integration of epigenome data with transcriptome and other epigenetic data, such as histone modifications, is important, since small DNA methylation changes might reflect larger changes in chromatin structure and could be associated with broader changes in gene expression. The genomic context should also be considered; small changes within a regulatory element such as a promoter, enhancer, or insulator may have functional significance. In this regard, DMRs for obesity, as well as regions affected by prenatal famine exposure and meQTL for metabolic trait loci have been observed to overlap enhancer elements [8, 43, 68]. There is evidence that DNA methylation in famine-associated regions could indeed affect enhancer activity [68], supporting a role of nutrition-induced methylation changes in gene regulation.

A major limitation in many human studies is that epigenetic marks are often assessed in peripheral blood, rather than in metabolically relevant tissues (Fig. 2). The heterogeneity of blood is an issue, since different cell populations have distinct epigenetic signatures, but algorithms have been developed to estimate the cellular composition to overcome this problem [86]. Perhaps more importantly, epigenetic marks in blood cells may not necessarily report the status of the tissues of primary interest. Despite this, recent studies have provided clear evidence of a relationship between epigenetic marks in blood cells and BMI. In the case of *HIF3A* for which the level of methylation (beta-value) in the study population ranged from 0.14–0.52, a 10 % increase in methylation was associated with a BMI increase of 7.8 %

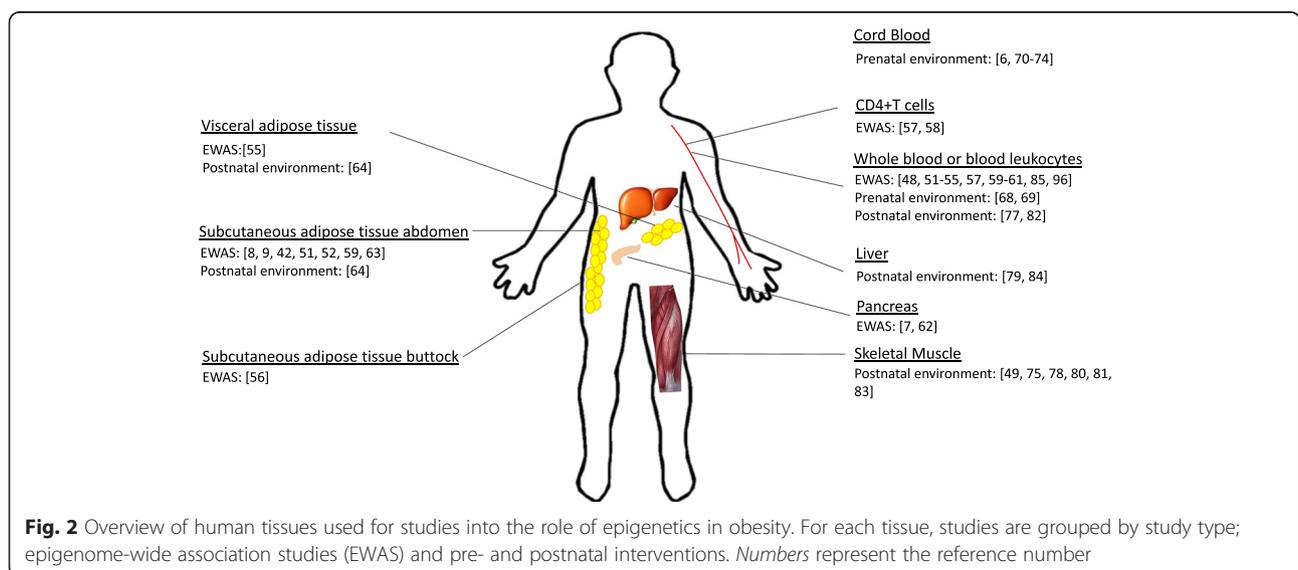
[52]. Likewise, a 10 % difference in *PGC1A* methylation may predict up to 12 % difference in fat mass [54].

Conclusions

The study of the role of epigenetics in obesity and metabolic disease has expanded rapidly in recent years, and evidence is accumulating of a link between epigenetic modifications and metabolic health outcomes in humans. Potential epigenetic biomarkers associated with obesity and metabolic health have also emerged from recent studies. The validation of epigenetic marks in multiple cohorts, the fact that several marks are found in genes with a plausible function in obesity and T2DM development, as well as the overlap of epigenetic marks with known obesity and T2DM genetic loci strengthens the evidence that these associations are real. Causality has so far been difficult to establish; however, regardless of whether the associations are causal, the identified epigenetic marks may still be relevant as biomarkers for obesity and metabolic disease risk.

Effect sizes in easily accessible tissues such as blood are small but do seem reproducible despite variation in ethnicity, tissue type, and analysis methods [51]. Also, even small DNA methylation changes may have biological significance. An integrative “omics” approach will be crucial in further unraveling the complex interactions between the epigenome, transcriptome, genome, and metabolic health. Longitudinal studies, ideally spanning multiple generations, are essential to establishing causal relationships. We can expect more such studies in the future, but this will take time.

While animal studies continue to demonstrate an effect of early life nutritional exposure on the epigenome and metabolic health of the offspring, human data are still limited. However, recent studies have provided clear



evidence that exposure to suboptimal nutrition during specific periods of prenatal development is associated with methylation changes in the offspring and therefore have the potential to influence adult phenotype. Animal studies will be important to verify human findings in a more controlled setting, help determine whether the identified methylation changes have any impact on metabolic health, and unravel the mechanisms underlying this intergenerational/transgenerational epigenetic regulation. The identification of causal mechanisms underlying metabolic memory responses, the mode of transmission of the phenotypic effects into successive generations, the degree of impact and stability of the transmitted trait, and the identification of an overarching and unifying evolutionary context also remain important questions to be addressed. The latter is often encapsulated by the predictive adaptive response hypothesis, i.e., a response to a future anticipated environment that increases fitness of the population. However, this hypothesis has increasingly been questioned as there is limited evidence for increased fitness later in life [87].

In summary, outcomes are promising, as the epigenetic changes are linked with adult metabolic health and they act as a mediator between altered prenatal nutrition and subsequent increased risk of poor metabolic health outcomes. New epigenetic marks have been identified that are associated with measures of metabolic health. Integration of different layers of genomic information has added further support to causal relationships, and there have been further studies showing effects of pre- and postnatal environment on the epigenome and health. While many important questions remain, recent methodological advances have enabled the types of large-scale population-based studies that will be required to address the knowledge gaps. The next decade promises to be a period of major activity in this important research area.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All authors contributed to the drafting and critical revision of the manuscript, and all authors read and approved the final manuscript.

Authors' information

Beverly S. Muhlhauser and Peter L. Molloy are joint last authors.

Acknowledgements

This work has been supported by a grant from the Science and Industry Endowment Fund (Grant RP03-064). JLM and BSM are supported by the National Health and Medical Research Council Career Development Fellowships (JLM, APP1066916; BSM, APP1004211). We thank Lance Macaulay and Sue Mitchell for critical reading and comments on the manuscript.

Author details

¹CSIRO Food and Nutrition Flagship, PO Box 52, North Ryde, NSW 1670, Australia. ²CSIRO Agriculture Flagship, 306 Carmody Road, St Lucia, QLD 4067, Australia. ³Early Origins of Adult Health Research Group, School of Pharmacy and Medical Sciences, Sansom Institute for Health Research, University of South Australia, GPO Box 2471, Adelaide, SA 5001, Australia.

⁴FOODplus Research Centre, Waite Campus, The University of Adelaide, PMB 1, Glen Osmond, SA 5064, Australia. ⁵Women's and Children's Health Research Institute, 72 King William Road, North Adelaide, SA 5006, Australia.

Received: 9 March 2015 Accepted: 29 June 2015

Published online: 11 July 2015

References

- WHO. WHO | Overweight and obesity. http://www.who.int/gho/ncd/risk_factors/overweight/en/index.html. Accessed 29 January 2015.
- Visscher PM, Brown MA, McCarthy MI, Yang J. Five years of GWAS discovery. *Am J Hum Genet.* 2012;90:7–24.
- Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature.* 2015;518:197–206.
- Ling C, Del Guerra S, Lupi R, Rönn T, Granhall C, Luthman H, et al. Epigenetic regulation of PPARGC1A in human type 2 diabetic islets and effect on insulin secretion. *Diabetologia.* 2008;51:615–22.
- Van Dijk SJ, Molloy PL, Varinli H, Morrison JL, Muhlhauser BS. Epigenetics and human obesity. *Int J Obes (Lond).* 2015;39:85–97.
- Teh AL, Pan H, Chen L, Ong M-L, Dogra S, Wong J, et al. The effect of genotype and in utero environment on interindividual variation in neonate DNA methylomes. *Genome Res.* 2014;24:1064–74.
- Olsson AH, Volkov P, Bacos K, Dayeh T, Hall E, Nilsson EA, et al. Genome-wide associations between genetic and epigenetic variation influence mRNA expression and insulin secretion in human pancreatic islets. *PLoS Genet.* 2014;10:e1004735.
- Grundberg E, Meduri E, Sandling JK, Hedman AK, Keildson S, Buil A, et al. Global analysis of DNA methylation variation in adipose tissue from twins reveals links to disease-associated variants in distal regulatory elements. *Am J Hum Genet.* 2013;93:876–90.
- Ronn T, Volkov P, Gillberg L, Kokosar M, Perflyev A, Jacobsen AL, et al. Impact of age, BMI and HbA1c levels on the genome-wide DNA methylation and mRNA expression patterns in human adipose tissue and identification of epigenetic biomarkers in blood. *Hum Mol Genet.* 2015;24:3792–813.
- Waterland RA, Michels KB. Epigenetic epidemiology of the developmental origins hypothesis. *Annu Rev Nutr.* 2007;27:363–88.
- McMillen IC, Rattanatray L, Duffield JA, Morrison JL, MacLaughlin SM, Gentili S, et al. The early origins of later obesity: pathways and mechanisms. *Adv Exp Med Biol.* 2009;646:71–81.
- Ravelli A, van der Meulen J, Michels R, Osmond C, Barker D, Hales C, et al. Glucose tolerance in adults after prenatal exposure to famine. *Lancet.* 1998;351:173–7.
- McMillen IC, MacLaughlin SM, Muhlhauser BS, Gentili S, Duffield JL, Morrison JL. Developmental origins of adult health and disease: the role of periconceptual and foetal nutrition. *Basic Clin Pharmacol Toxicol.* 2008;102:82–9.
- Zhang S, Rattanatray L, McMillen IC, Suter CM, Morrison JL. Periconceptual nutrition and the early programming of a life of obesity or adversity. *Prog Biophys Mol Biol.* 2011;106:307–14.
- Bouret S, Levin BE, Ozanne SE. Gene-environment interactions controlling energy and glucose homeostasis and the developmental origins of obesity. *Physiol Rev.* 2015;95:47–82.
- Borengasser SJ, Zhong Y, Kang P, Lindsey F, Ronis MJ, Badger TM, et al. Maternal obesity enhances white adipose tissue differentiation and alters genome-scale DNA methylation in male rat offspring. *Endocrinology.* 2013;154:4113–25.
- Gluckman PD, Lillycrop KA, Vickers MH, Pleasants AB, Phillips ES, Beedle AS, et al. Metabolic plasticity during mammalian development is directionally dependent on early nutritional status. *Proc Natl Acad Sci U S A.* 2007;104:12796–800.
- Godfrey KM, Sheppard A, Gluckman PD, Lillycrop KA, Burdge GC, McLean C, et al. Epigenetic gene promoter methylation at birth is associated with child's later adiposity. *Diabetes.* 2011;60:1528–34.
- McMillen IC, Adam CL, Muhlhauser BS. Early origins of obesity: programming the appetite regulatory system. *J Physiol.* 2005;565(Pt 1):9–17.
- Begum G, Stevens A, Smith EB, Connor K, Challis JR, Bloomfield F, et al. Epigenetic changes in fetal hypothalamic energy regulating pathways are associated with maternal undernutrition and twinning. *FASEB J.* 2012;26:1694–703.

21. Ge ZJ, Liang QX, Hou Y, Han ZM, Schatten H, Sun QY, et al. Maternal obesity and diabetes may cause DNA methylation alteration in the spermatozoa of offspring in mice. *Reprod Biol Endocrinol*. 2014;12:29.
22. Jousse C, Parry L, Lambert-Langlais S, Maurin AC, Averous J, Bruhat A, et al. Perinatal undernutrition affects the methylation and expression of the leptin gene in adults: implication for the understanding of metabolic syndrome. *FASEB J*. 2011;25:3271–8.
23. Lan X, Cretney EC, Kropp J, Khateeb K, Berg MA, Penagaricano F, et al. Maternal diet during pregnancy induces gene expression and DNA methylation changes in fetal tissues in sheep. *Front Genet*. 2013;4:49.
24. Li CC, Young PE, Maloney CA, Eaton SA, Cowley MJ, Buckland ME, et al. Maternal obesity and diabetes induces latent metabolic defects and widespread epigenetic changes in isogenic mice. *Epigenetics*. 2013;8:602–11.
25. Lillycrop KA, Phillips ES, Jackson AA, Hanson MA, Burdge GC. Dietary protein restriction of pregnant rats induces and folic acid supplementation prevents epigenetic modification of hepatic gene expression in the offspring. *J Nutr*. 2005;135:1382–6.
26. Radford EJ, Ito M, Shi H, Corish JA, Yamazawa K, Isganaitis E, et al. In utero effects. In utero undernourishment perturbs the adult sperm methylome and intergenerational metabolism. *Science*. 2014;345(80):1255–903.
27. Suter M, Bockock P, Showalter L, Hu M, Shope C, McKnight R, et al. Epigenomics: maternal high-fat diet exposure in utero disrupts peripheral circadian gene expression in nonhuman primates. *FASEB J*. 2011;25:714–26.
28. Suter MA, Ma J, Vuguin PM, Hartil K, Fiallo A, Harris RA, et al. In utero exposure to a maternal high-fat diet alters the epigenetic histone code in a murine model. *Am J Obs Gynecol*. 2014;210:463 e1–463 e11.
29. Tosh DN, Fu Q, Callaway CW, McKnight RA, McMillen IC, Ross MG, et al. Epigenetics of programmed obesity: alteration in IUGR rat hepatic IGF1 mRNA expression and histone structure in rapid vs. delayed postnatal catch-up growth. *Am J Physiol Gastrointest Liver Physiol*. 2010;299:G1023–9.
30. Sandovici I, Smith NH, Nitert MD, Ackers-Johnson M, Uribe-Lewis S, Ito Y, et al. Maternal diet and aging alter the epigenetic control of a promoter-enhancer interaction at the *Hnf4a* gene in rat pancreatic islets. *Proc Natl Acad Sci U S A*. 2011;108:5449–54.
31. Braunschweig M, Jagannathan V, Gutzwiller A, Bee G. Investigations on transgenerational epigenetic response down the male line in F2 pigs. *PLoS One*. 2012;7, e30583.
32. Carone BR, Fauquier L, Habib N, Shea JM, Hart CE, Li R, et al. Paternally induced transgenerational environmental reprogramming of metabolic gene expression in mammals. *Cell*. 2010;143:1084–96.
33. Ost A, Lempradl A, Casas E, Weigert M, Tiko T, Deniz M, et al. Paternal diet defines offspring chromatin state and intergenerational obesity. *Cell*. 2014;159:1352–64.
34. Martínez D, Pentinat T, Ribó S, Daviaud C, Bloks VW, Cebrià J, et al. In utero undernutrition in male mice programs liver lipid metabolism in the second-generation offspring involving altered *Lxra* DNA methylation. *Cell Metab*. 2014;19:941–51.
35. Wei Y, Yang C-R, Wei Y-P, Zhao Z-A, Hou Y, Schatten H, et al. Paternally induced transgenerational inheritance of susceptibility to diabetes in mammals. *Proc Natl Acad Sci U S A*. 2014;111:1873–8.
36. Grossniklaus U, Kelly WG, Kelly B, Ferguson-Smith AC, Pembrey M, Lindquist S. Transgenerational epigenetic inheritance: how important is it? *Nat Rev Genet*. 2013;14:228–35.
37. Pembrey M, Saffery R, Bygren LO. Human transgenerational responses to early-life experience: potential impact on development, health and biomedical research. *J Med Genet*. 2014;51:563–72.
38. Wolff GL, Kodell RL, Moore SR, Cooney CA. Maternal epigenetics and methyl supplements affect agouti gene expression in *Avy/a* mice. *FASEB J*. 1998;12:949–57.
39. Jirtle RL, Skinner MK. Environmental epigenomics and disease susceptibility. *Nat Rev Genet*. 2007;8:253–62.
40. Morgan HD, Sutherland HG, Martin DI, Whitelaw E. Epigenetic inheritance at the agouti locus in the mouse. *Nat Genet*. 1999;23:314–8.
41. Cropley JE, Suter CM, Beckman KB, Martin DI. Germ-line epigenetic modification of the murine *Avy* allele by nutritional supplementation. *Proc Natl Acad Sci U S A*. 2006;103:17308–12.
42. Hoile SP, Lillycrop KA, Thomas NA, Hanson MA, Burdge GC. Dietary protein restriction during F0 pregnancy in rats induces transgenerational changes in the hepatic transcriptome in female offspring. *PLoS One*. 2011;6, e21668.
43. Multhaup ML, Seldin MM, Jaffe AE, Lei X, Kirchner H, Mondal P, et al. Mouse-human experimental epigenetic analysis unmasks dietary targets and genetic liability for diabetic phenotypes. *Cell Metab*. 2015;21:138–49.
44. Michels KB, Binder AM, Dedeurwaerder S, Epstein CB, Grealley JM, Gut I, et al. Recommendations for the design and analysis of epigenome-wide association studies. *Nat Methods*. 2013;10:949–55.
45. Dayeh TA, Olsson AH, Volkov P, Almgren P, Rönn T, Ling C. Identification of CpG-SNPs associated with type 2 diabetes and differential DNA methylation in human pancreatic islets. *Diabetologia*. 2013;56:1036–46.
46. Relton CL, Davey Smith G. Two-step epigenetic Mendelian randomization: a strategy for establishing the causal role of epigenetic processes in pathways to disease. *Int J Epidemiol*. 2012;41:161–76.
47. Liu Y, Aryee MJ, Padyukov L, Fallin MD, Hesselberg E, Runarsson A, et al. Epigenome-wide association data implicate DNA methylation as an intermediary of genetic risk in rheumatoid arthritis. *Nat Biotechnol*. 2013;31:142–7.
48. Yuan W, Xia Y, Bell CG, Yet I, Ferreira T, Ward KJ, et al. An integrated epigenomic analysis for type 2 diabetes susceptibility loci in monozygotic twins. *Nat Commun*. 2014;5:5719.
49. Nitert MD, Dayeh T, Volkov P, Elgzyri T, Hall E, Nilsson E, et al. Impact of an exercise intervention on DNA methylation in skeletal muscle from first-degree relatives of patients with type 2 diabetes. *Diabetes*. 2012;61:3322–32.
50. Gagnon F, Aïssi D, Carrié A, Morange P-E, Tréguouët D-A. Robust validation of methylation levels association at *CPT1A* locus with lipid plasma levels. *J Lipid Res*. 2014;55:1189–91.
51. Demerath EW, Guan W, Grove ML, Aslibekyan S, Mendelson M, Zhou Y-H, et al. Epigenome-wide association study (EWAS) of BMI, BMI change, and waist circumference in African American adults identifies multiple replicated loci. *Hum Mol Genet*. 2015;ddv161–.
52. Dick KJ, Nelson CP, Tsaprouni L, Sandling JK, Aïssi D, Wahl S, et al. DNA methylation and body-mass index: a genome-wide analysis. *Lancet*. 2014;6736:1–9.
53. Su S, Zhu H, Xu X, Wang X, Dong Y, Kapuku G, et al. DNA methylation of the *LY86* gene is associated with obesity, insulin resistance, and inflammation. *Twin Res Hum Genet*. 2014;17:183–91.
54. Clarke-Harris R, Wilkin TJ, Hosking J, Pinkney J, Jeffery AN, Metcalf BS, et al. *PGC1α* promoter methylation in blood at 5–7 years predicts adiposity from 9 to 14 years (EarlyBird 50). *Diabetes*. 2014;63:2528–37.
55. Guay S-P, Brisson D, Lamarche B, Biron S, Lesclercq O, Biertho L, et al. *ADRB3* gene promoter DNA methylation in blood and visceral adipose tissue is associated with metabolic disturbances in men. *Epigenomics*. 2014;6:33–43.
56. Agha G, Houseman EA, Kelsey KT, Eaton CB, Buka SL, Loucks EB. Adiposity is associated with DNA methylation profile in adipose tissue. *Int J Epidemiol*. 2014:1–11.
57. Irvin MR, Zhi D, Joehanes R, Mendelson M, Aslibekyan S, Claas SA, et al. Epigenome-wide association study of fasting blood lipids in the genetics of lipid-lowering drugs and diet network study. *Circulation*. 2014;130:565–72.
58. Frazier-Wood AC, Aslibekyan S, Absher DM, Hopkins PN, Sha J, Tsai MY, et al. Methylation at *CPT1A* locus is associated with lipoprotein subfraction profiles. *J Lipid Res*. 2014;55:1324–30.
59. Pfeifferm L, Wahl S, Pilling LC, Reischl E, Sandling JK, Kunze S, et al. DNA methylation of lipid-related genes affects blood lipid levels. *Circ Cardiovasc Genet*. 2015.
60. Petersen A-K, Zeilinger S, Kastenmüller G, Römisch-Margl W, Brügger M, Peters A, et al. Epigenetics meets metabolomics: an epigenome-wide association study with blood serum metabolic traits. *Hum Mol Genet*. 2014;23:534–45.
61. Hidalgo B, Irvin MR, Sha J, Zhi D, Aslibekyan S, Absher D, et al. Epigenome-wide association study of fasting measures of glucose, insulin, and HOMA-IR in the genetics of lipid lowering drugs and diet network study. *Diabetes*. 2014;63:801–7.
62. Dayeh T, Volkov P, Salö S, Hall E, Nilsson E, Olsson AH, et al. Genome-wide DNA methylation analysis of human pancreatic islets from type 2 diabetic and non-diabetic donors identifies candidate genes that influence insulin secretion. *PLoS Genet*. 2014;10, e1004160.
63. Nilsson E, Jansson PA, Perflyev A, Volkov P, Pedersen M, Svensson MK, et al. Altered DNA methylation and differential expression of genes influencing metabolism and inflammation in adipose tissue from subjects with type 2 diabetes. *Diabetes*. 2014;63:2962–76.
64. Benton MC, Johnstone A, Eccles D, Harmon B, Hayes MT, Lea RA, et al. An analysis of DNA methylation in human adipose tissue reveals differential

- modification of obesity genes before and after gastric bypass and weight loss. *Gene*. 2015;16:1–21.
65. Bateson P, Gluckman P. Plasticity and robustness in development and evolution. *Int J Epidemiol*. 2012;41:219–23.
 66. Feinberg AP, Irizarry RA, Feinberg AP, Irizarry RA. Evolution in health and medicine Sackler colloquium: stochastic epigenetic variation as a driving force of development, evolutionary adaptation, and disease. *Proc Natl Acad Sci U S A*. 2010;107(Suppl):1757–64.
 67. Martino D, Loke YJ, Gordon L, Ollikainen M, Cruickshank MN, Saffery R, et al. Longitudinal, genome-scale analysis of DNA methylation in twins from birth to 18 months of age reveals rapid epigenetic change in early life and pair-specific effects of discordance. *Genome Biol*. 2013;14:R42.
 68. Tobi EW, Goeman JJ, Monajemi R, Gu H, Putter H, Zhang Y, et al. DNA methylation signatures link prenatal famine exposure to growth and metabolism. *Nat Commun*. 2014;5:5592.
 69. Dominguez-Salas P, Moore SE, Baker MS, Bergen AW, Cox SE, Dyer RA, et al. Maternal nutrition at conception modulates DNA methylation of human metastable epialleles. *Nat Commun*. 2014;5:3746.
 70. Quilter CR, Cooper WN, Cliffe KM, Skinner BM, Prentice PM, Nelson L, et al. Impact on offspring methylation patterns of maternal gestational diabetes mellitus and intrauterine growth restraint suggest common genes and pathways linked to subsequent type 2 diabetes risk. *FASEB J*. 2014:1–12.
 71. Morales E, Groom A, Lawlor DA, Relton CL. DNA methylation signatures in cord blood associated with maternal gestational weight gain: results from the ALSPAC cohort. *BMC Res Notes*. 2014;7:278.
 72. Ruchat SM, Houde AA, Voisin G, St-Pierre J, Perron P, Baillargeon JP, et al. Gestational diabetes mellitus epigenetically affects genes predominantly involved in metabolic diseases. *Epigenetics*. 2013;8:935–43.
 73. Liu X, Chen Q, Tsai H-J, Wang G, Hong X, Zhou Y, et al. Maternal preconception body mass index and offspring cord blood DNA methylation: exploration of early life origins of disease. *Environ Mol Mutagen*. 2014;55:223–30.
 74. Soubry A, Murphy SK, Wang F, Huang Z, Vidal AC, Fuemmeler BF, et al. Newborns of obese parents have altered DNA methylation patterns at imprinted genes. *Int J Obes (Lond)*. 2015;39:650–7.
 75. Jacobsen SC, Brøns C, Bork-Jensen J, Ribel-Madsen R, Yang B, Lara E, et al. Effects of short-term high-fat overfeeding on genome-wide DNA methylation in the skeletal muscle of healthy young men. *Diabetologia*. 2012;55:3341–9.
 76. Gillberg L, Jacobsen SC, Rönn T, Brøns C, Vaag A. PPARGC1A DNA methylation in subcutaneous adipose tissue in low birth weight subjects—impact of 5 days of high-fat overfeeding. *Metabolism*. 2014;63:263–71.
 77. Huang Y-T, Maccani JZJ, Hawley NL, Wing RR, Kelsey KT, McCaffery JM. Epigenetic patterns in successful weight loss maintainers: a pilot study. *Int J Obes (Lond)*. 2015;39:865–8.
 78. Barres R, Kirchner H, Rasmussen M, Yan J, Kantor FR, Krook A, Näslund E, Zierath JR. Weight loss after gastric bypass surgery in human obesity remodels promoter methylation. *Cell Rep*. 2013:1–8.
 79. Ahrens M, Ammerpohl O, von Schönfels W, Kolarova J, Bens S, Itzel T, et al. DNA methylation analysis in nonalcoholic fatty liver disease suggests distinct disease-specific and remodeling signatures after bariatric surgery. *Cell Metab*. 2013;18:296–302.
 80. Voisin S, Eynon N, Yan X, Bishop DJ. Exercise training and DNA methylation in humans. *Acta Physiol (Oxf)*. 2014;213:39–59.
 81. Lindholm ME, Marabita F, Gomez-Cabrero D, Rundqvist H, Ekström TJ, Tegnér J, et al. An integrative analysis reveals coordinated reprogramming of the epigenome and the transcriptome in human skeletal muscle after training. *Epigenetics*. 2014;9:1557–69.
 82. Denham J, O'Brien BJ, Marques FZ, Charchar FJ. Changes in the leukocyte methylome and its effect on cardiovascular related genes after exercise. *J Appl Physiol*. 2014;jap.00878.2014.
 83. Rowlands DS, Page RA, Sukala WR, Giri M, Ghimbovschi SD, Hayat I, et al. Multi-omic integrated networks connect DNA methylation and miRNA with skeletal muscle plasticity to chronic exercise in type 2 diabetic obesity. *Physiol Genomics*. 2014;46:747–65.
 84. Horvath S, Erhart W, Brosch M, Ammerpohl O, von Schönfels W, Ahrens M, et al. Obesity accelerates epigenetic aging of human liver. *Proc Natl Acad Sci*. 2014;111:15538–43.
 85. Almén MS, Nilsson EK, Jacobsson JA, Kalnina I, Klovins J, Fredriksson R, et al. Genome-wide analysis reveals DNA methylation markers that vary with both age and obesity. *Gene*. 2014;548:61–7.
 86. Houseman EA, Molitor J, Marsit CJ. Reference-free cell mixture adjustments in analysis of DNA methylation data. *Bioinformatics*. 2014;30:1431–9.
 87. Wells JC. A critical appraisal of the predictive adaptive response hypothesis. *Int J Epidemiol*. 2012;41:229–35.
 88. Williams-Wyss O, Zhang S, MacLaughlin SM, Kleemann D, Walker SK, Suter CM, et al. Embryo number and periconceptual undernutrition in the sheep have differential effects on adrenal epigenotype, growth, and development. *Am J Physiol Endocrinol Metab*. 2014;307:E141–50.
 89. Zhang S, Rattanaray L, Morrison JL, Nicholas LM, Lie S, McMillen IC. Maternal obesity and the early origins of childhood obesity: weighing up the benefits and costs of maternal weight loss in the periconceptual period for the offspring. *Exp Diabetes Res*. 2011;2011:585749.
 90. Zhang S, Williams-Wyss O, MacLaughlin SM, Walker SK, Kleemann DO, Suter CM, et al. Maternal undernutrition during the first week after conception results in decreased expression of glucocorticoid receptor mRNA in the absence of GR exon 17 hypermethylation in the fetal pituitary in late gestation. *J Dev Orig Heal Dis*. 2013;4:391–401.
 91. Lie S, Morrison JL, Williams-Wyss O, Suter CM, Humphreys DT, Ozanne SE, et al. Periconceptual undernutrition programs changes in insulin-signaling molecules and microRNAs in skeletal muscle in singleton and twin fetal sheep. *Biol Reprod*. 2014;90:5.
 92. Van Straten EM, van Meer H, Huijckman NC, van Dijk TH, Baller JF, Verkade HJ, et al. Fetal liver X receptor activation acutely induces lipogenesis but does not affect plasma lipid response to a high-fat diet in adult mice. *Am J Physiol Endocrinol Metab*. 2009;297:E1171–8.
 93. Fernandez-Twinn DS, Alfaradhi MZ, Martin-Gronert MS, Duque-Guimaraes DE, Piekarz A, Ferland-McCollough D, et al. Downregulation of IRS-1 in adipose tissue of offspring of obese mice is programmed cell-autonomously through post-transcriptional mechanisms. *Mol Metab*. 2014;3:325–33.
 94. Waterland RA, Travisano M, Tahiliani KG. Diet-induced hypermethylation at agouti viable yellow is not inherited transgenerationally through the female. *FASEB J*. 2007;21:3380–5.
 95. Ge ZJ, Luo SM, Lin F, Liang QX, Huang L, Wei YC, et al. DNA methylation in oocytes and liver of female mice and their offspring: effects of high-fat-diet-induced obesity. *Env Heal Perspect*. 2014;122:159–64.
 96. Ollikainen M, Ismail K, Gervin K, Kyllönen A, Hakkarainen A, Lundbom J, et al. Genome-wide blood DNA methylation alterations at regulatory elements and heterochromatic regions in monozygotic twins discordant for obesity and liver fat. *Clin Epigenetics*. 2015;7:1–13.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

